1 Figure S1: Genital tract histopathology at week 12. Genital tract tissues obtained from 2 animals in Group 1 (black squares), Group 2 (white triangles), and uninfected controls (UI, black 3 diamonds) at tissue harvest were analyzed histologically. The highest levels of inflammation 4 were detected in the endocervix (A) of animals in Group 1. Inflammation was similar between 5 groups in the uterus (B), oviducts (C), and fimbriae (D). Symbols represent individual animals 6 (A, B) or oviducts (C, D). \* P < 0.05 by One way ANOVA with Dunn's Multiple Comparison test. 7 8 Figure S2: Chlamydia-specific antibody in the serum and secretions was similar between 9 animals in Group 1 and Group 2. Levels of IgG in the serum (A), IgA in the serum (B), IgG in 10 the secretions (C), and IgA in the secretions (D) did not differ at week 6 or week 12 for animals 11 in Group 1 (black bars) or Group 2 (white bars). P > 0.05 by One way ANOVA with Dunn's 12 Multiple Comparison test. Graphs show the log<sub>2</sub> of the largest antibody dilution where antibody 13 to elementary bodies was detected at a level above the negative control (-). Origin of the y-axis 14 is set at the lowest dilution of the negative control sample. Numbers on the x-axis indicate the 15 animal number. 16 17 Figure S3: Representative flow plot depicting gating strategy used to define proliferative 18 T cells. Dead cells were excluded using Live/Dead stain, and (A) CD3+ CD4+ or (B) 19 CD3+CD8+ T cells were analyzed for expression of Ki67, a marker that is upregulated in 20 proliferating cells. 21 22 Figure S4: Chlamydia-specific responses were similar in the peripheral blood of animals 23 in Group 1 and Group 2. Proliferation of (A) CD3+CD4+ T cells and (B) CD3+ CD8+ T cells 24 was determined by analysis of Ki67 expression by flow cytometry after 5 days of incubation with 25 EBs. Levels of IL-2 (C) and IFNy (D) were measured in the supernatants with significantly 26 higher levels of IFNy detected in the supernatants of animals in Group 1 Controllers (black

squares, dashed line) relative to animals in Group 2 (white triangles, solid line) at week 12. Data points indicate values obtained for PBMCs incubated with EBs for 5 days after subtraction of values for PBMCs incubated without EBs. Line indicates mean. \*P < 0.05 by Two-way RM with Bonferroni posttests. Figure S5. Representative flow plot depicting gating strategy used for populations of memory T cells. Dead cells were excluded using Live/Dead stain, and (A) CD3+ CD4+ or (B) CD3+CD8+ T cells were subdivided into CD28<sup>+</sup>CD95<sup>+</sup> central memory T cells (T<sub>CM</sub>) and CD28<sup>-</sup> CD95<sup>+</sup> effector memory T cells (T<sub>EM</sub>). Figure S6. Comparison of effector and central memory T cells in the peripheral blood of animals in Group 1 and Group 2. Frequencies of Chlamydia-specific CD4+ effector memory cells (A), CD4+ central memory cells (B), CD8+ effector memory cells (C), and CD8+ central memory cells (D) were similar in the peripheral blood of animals in Group 1 (black circles, dashed line) and Group 2 (white squares, solid line). T<sub>CM</sub>: CD28<sup>+</sup>CD95<sup>+</sup>. T<sub>EM</sub>: CD28<sup>-</sup>CD95<sup>+</sup>. Data points indicate frequencies obtained for PBMCs incubated with EBs for 5 days after values for PBMCs incubated without EBs were subtracted. Line indicates mean.

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Fig. S1

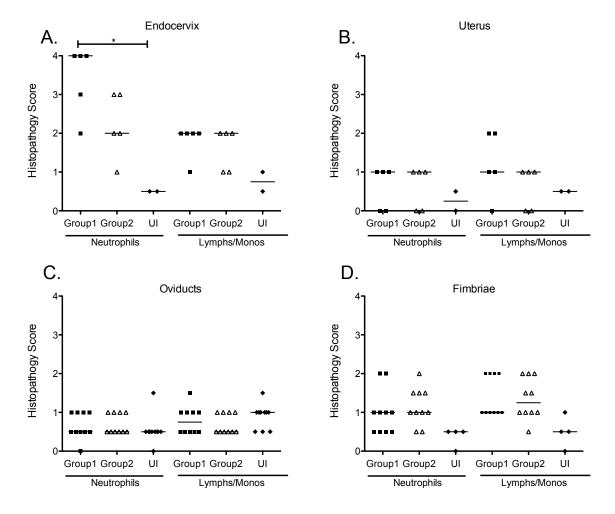


Fig. S2

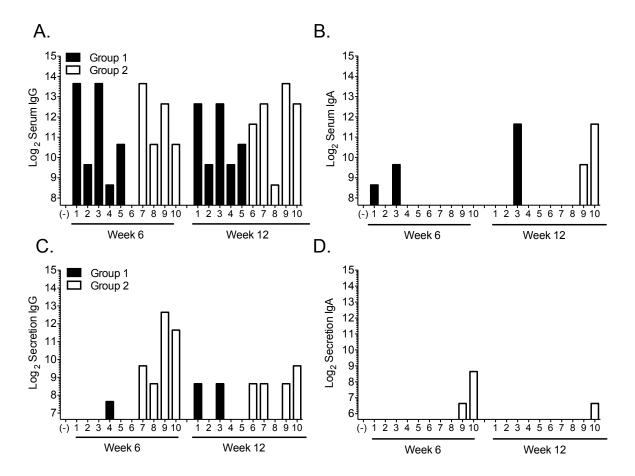


Fig. S3

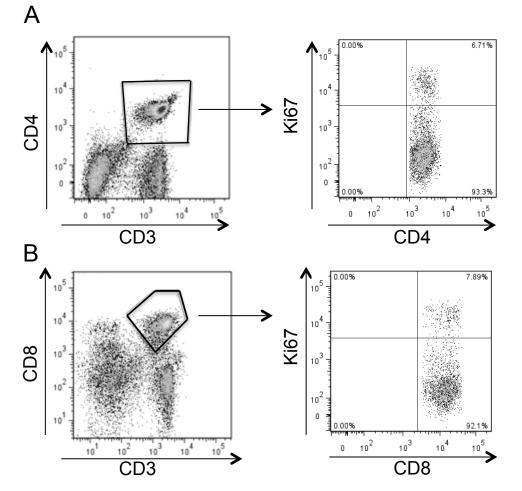


Fig. S4

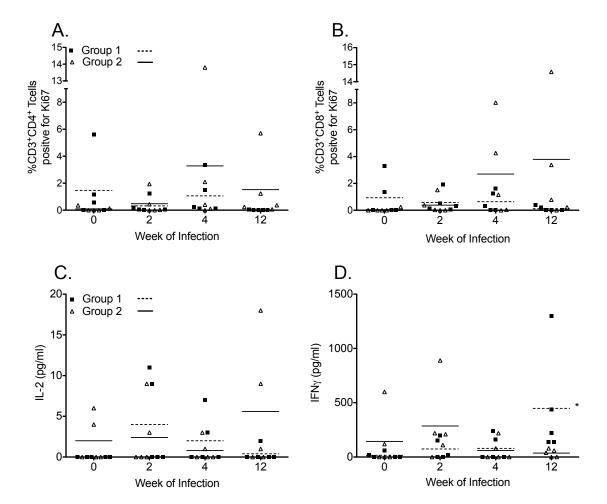


Fig. S5

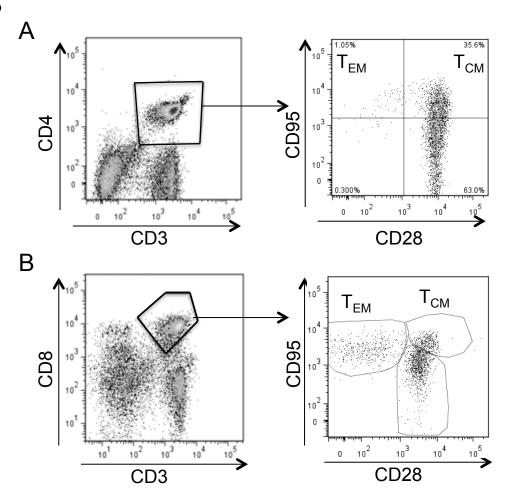


Fig. S6

